I. Background

Investigators may need to identify frogs, specifically *Xenopus spp.*, producing oocytes of sufficient quality for the proposed experiments. For example, oocytes are sometimes used for the transcription and/or translation of exogenous DNA or RNA templates. Oocytes from different frogs have been observed to have very different efficiencies of DNA transcription or RNA translation. The quality of oocytes from any given frog is usually constant over several months, so that a frog that is found to be of good quality (i.e., one that produces oocytes showing acceptable quality in the relevant transcriptional or translational assays) can be re-used with an increased assurance of high quality oocytes. Therefore, investigators may need to perform multiple partial ovariectomies in *Xenopus spp.* (MPOX).

II. Policy

Investigators may perform multiple partial ovariectomies on *Xenopus spp.* if there is a clear and compelling scientific rationale for why multiple partial ovariectomies in *Xenopus* (MPOX) are justified for their research.

III. Responsibilities

A. Animal Care and Use Committee (ACUC) – MPOX must be scientifically justified in the animal use protocol and approved by the ACUC. Tracking procedures and record keeping should be considered at semi-annual inspections.

B. Office of Laboratory Animal Care (OLAC) – OLAC animal care personnel should ask for clarification from the lab if tracking procedures are not clear.

C. Principal Investigator (PI) – The PI is responsible for ensuring that all lab personnel performing MPOX or overseeing frogs that have undergone MPOX understand and abide by this policy.

IV. Required Procedures

A. A description of the surgeries and related procedures as well as a clear and compelling scientific rationale for why MPOX are justified for their research must be included in the PI’s Animal Use Protocol (AUP). Conservation of animal numbers (except for endangered species), economy of animal husbandry, cost of animals, and similar considerations are not acceptable rationale.

B. Maximum number of surgeries and inter-surgical intervals
1. No more than four (4) partial ovariectomies may be performed per frog, with a minimum interval between surgeries of no less than three (3) weeks.

2. The fourth and final ovariectomy must be a terminal surgery.

C. Tracking and Record Keeping

1. If the rationale for performing MPOX is to identify and re-use individual frogs known to produce optimal oocytes, the investigator must provide a plan for:
   a) Tracking these frogs;
   b) Tracking the number of surgeries performed on each frog, and;
   c) Ensuring the minimum interval of three (3) weeks between surgeries.

2. Procedures for tracking can include, but are not limited to, segregation of operated animals into different post-surgical holding tanks, or physical marking of individual animals. Physical identification may include, but is not limited to, freeze branding, tattooing, color coded beads, photography of the unique markings on each animals' dorsum, implantation of sterile electronic personal information transponders (PIT), or elastomer tags. The identification method must be described in the approved AUP.

3. Investigators must document the number of surgeries performed, the dates upon which and by whom they were performed, and the postsurgical monitoring for frogs subjected to MPOX. The methods used to document the number of surgeries must be described in the AUP. These records must be readily available for review during inspections of the animals and/or their housing area by representatives from OLAC, the ACUC, or external regulatory or accrediting agencies.

D. Surgery and Anesthesia

1. All MPOX must be performed aseptically and in accordance with campus surgery guidelines. The surgical site must be gently swabbed with sterile saline or other appropriate mild disinfectant (e.g. 0.75% chlorhexidine).

2. Frogs must be adequately anesthetized prior to surgery with an anesthetic regimen described in the AUP. Anesthetic depth must be checked by toe pinch every 15 minutes and prior to start of surgery. This must be recorded in the anesthetic record. Please refer to ACUC Recordkeeping Guidelines for Surgical
Procedures on Laboratory Animals for additional information.

3. Hypothermia is not an acceptable method of immobilization (2013 AVMA Guidelines for Euthanasia). Examples of acceptable anesthetics include pharmaceutical grade solutions of benzocaine or buffered tricaine (MS-222). Anesthetic method must be recorded in the animal’s record.

4. Peri-operative Analgesia:
   a. The administration of analgesia is required and may be administered before surgery once the frog has been anesthetized, intra-operatively, or during recovery. Lack of analgesic use must be justified in the AUP. The following agents and doses are recommended.
   b. A local anesthetic, such as 0.25% bupivacaine is highly recommended to provide analgesia. Prior to closure by suture, the incision site should be swabbed with 0.25% bupivacaine, lidocaine, or other appropriate local anesthetic. Do not exceed 5 mg/kg total dose either topical or intra-incisional.
   c. In addition or alternatively the following analgesics may be utilized to provide systemic analgesia: flunixin meglumine, meloxicam, or butorphanol.

5. Nares must not be submerged during anesthesia and recovery. Skin should be kept moist throughout the procedure.

E. Post-operative Monitoring
   1. Frogs must be monitored by laboratory personnel daily for at least 48 hours. Frogs must then be monitored every 2-3 days for at least 2 weeks. Post-operative monitoring must be recorded in each animals' surgical record.
   2. Sutures must be removed in 2-3 weeks.
   3. Euthanasia of the animal must be recorded if animal is undergoing terminal surgery.

V. References

2013 AVMA Guidelines for Euthanasia (page 78 in particular)


Reference UNC: http://research.unc.edu/offices/iacuc/policies-procedures/